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THEME

**Identification of Hub genes and key biomarkers in patients with
breast cancer by bioinformatics tools**

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for giving us the strength and willpower to complete this work.*

الحمد لله

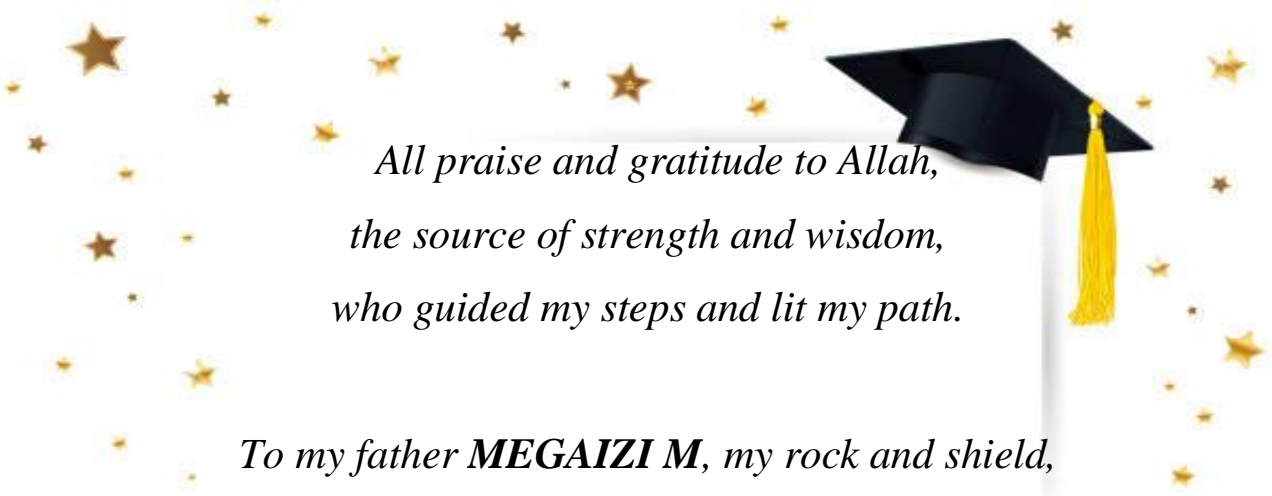
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Dedication



*All praise and gratitude to Allah,
the source of strength and wisdom,
who guided my steps and lit my path.*

*To my father **MEGAIZI M**, my rock and shield,
whose unwavering support gave me wings to rise.*

*To my dearest mother **BECHAKLIA FZ**, the heartbeat of
my soul, whose love and prayers shaped my journey.*

*To my brothers and sisters, the silent warriors behind my
success, always lifting me higher.*

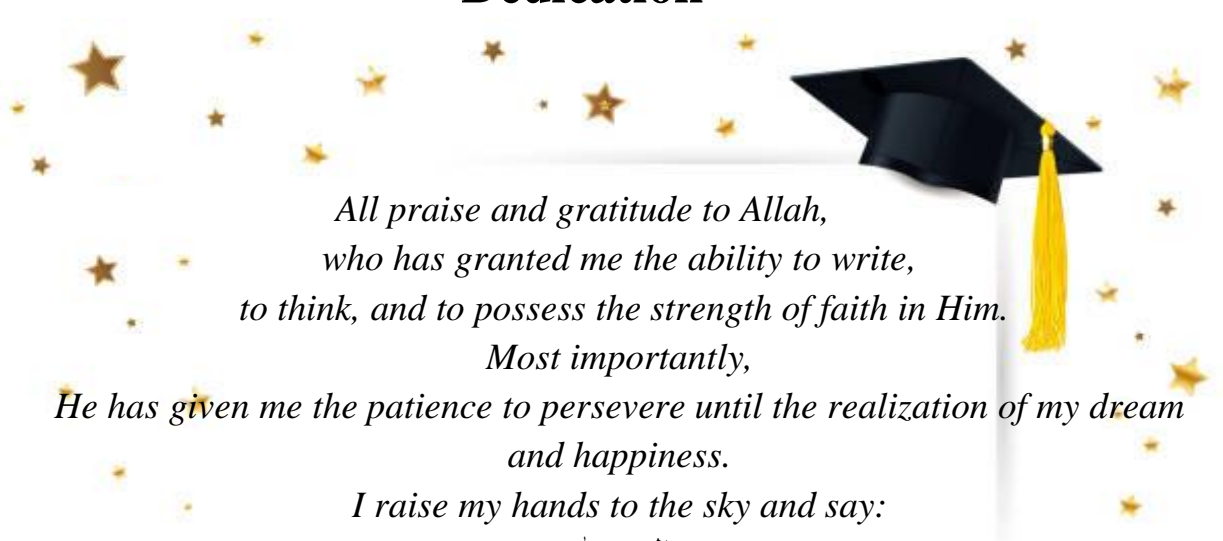
*And to every soul who lent a hand,
whispered a word of encouragement, or believed in me
this victory is not mine alone; it belongs to us all.*

*With gratitude,
with love,
with honor
I dedicate this work to you.*

Thank you!

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Dedication



*All praise and gratitude to Allah,
who has granted me the ability to write,
to think, and to possess the strength of faith in Him.*

*Most importantly,
He has given me the patience to persevere until the realization of my dream
and happiness.*

I raise my hands to the sky and say:

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carry with pride, and to the man who has always supported me:
my dear father **DJABALI N.***

*And to my angel in life, to the feeling of love, tenderness, and sincerity, to the
smile of life and the secret of existence, and to the one whose prayers were
the secret of my success:
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pillar and my first and last refuge, to the one who cleared the thorns of
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List of abbreviation

Abbreviation	Signification
ABUS	<i>Automated Breast Ultrasound</i>
AKT	<i>Protein Kinase B</i>
AMPK	<i>AMP-Activated Protein Kinase</i>
APC	<i>Anaphase-Promoting Complex</i>
APC/C	<i>Anaphase-Promoting Complex/Cyclosome</i>
BC	<i>Breast Cancer</i>
BP	<i>Biological Process</i>
BRCA1	<i>Breast Cancer Gene 1</i>
BRCA2	<i>Breast Cancer Gene 2</i>
BSE	<i>Breast Self-Exams</i>
CC	<i>Cellular Component</i>
CEM	<i>Contrast-Enhanced Mammography</i>
CIN	<i>Chromosomal Instability</i>
Cm	<i>Centimeter</i>
DAVID	<i>Database For Annotation, Visualization, And Integrated Discovery</i>
DBT	<i>Digital Breast Tomosynthesis</i>
DCIS	<i>Ductal Carcinoma in Situ</i>
DEGs	<i>Differentially Expressed Genes</i>
DNA	<i>Deoxyribonucleic Acid</i>
E2F1	<i>Erythroid 2 Factor 1</i>
ECM	<i>Extracellular Matrix</i>
EMT	<i>Epithelial-Mesenchymal Transition</i>
ER	<i>Estrogen Receptors</i>
FC	<i>Fold Change</i>
GCO	<i>Global Cancer Observatory</i>
GEO	<i>Gene Expression Omnibus</i>
GEPIA2	<i>Gene Expression Profiling Interactive Analysis 2</i>
GO	<i>Gene Ontology</i>
GPL	<i>Gene Platform</i>

GPSM2	<i>G-Protein Signaling Modulator 2</i>
GSE	<i>Gene Series Expression</i>
HER2	<i>Human Epidermal Growth Factor Receptor 2</i>
HR	<i>Hormone Receptors</i>
HR	<i>Hazard Ratios</i>
IHC	<i>Immunohistochemical</i>
KEGG	<i>Kyoto Encyclopedia of Genes and Genomes</i>
Ki-67	<i>Kiel 67</i>
LCIS	<i>Lobular Carcinoma in Situ</i>
MAM	<i>Mammography</i>
Mad1	<i>Mitotic Arrest Deficient 1</i>
Mad2	<i>Mitotic Arrest Deficiency Protein 2</i>
MAX	<i>Myc-associated Factor X</i>
MBC	<i>Metastatic Breast Cancer</i>
MCODE	<i>Molecular Complex Detection</i>
MF	<i>Molecular Function</i>
MiRNAs	<i>Micro Ribonucleic Acids</i>
MRI	<i>Magnetic Resonance Imaging</i>
mRNA	<i>Messenger Ribonucleic Acid</i>
MYC	<i>Myelocytomatosis</i>
OS	<i>Overall Survival</i>
P value	<i>Probability Value</i>
P21	<i>Protein 21</i>
P53	<i>Protein 53</i>
PI3K-Akt	<i>Phosphoinositide 3-Kinase</i>
PIK3CA	<i>Phosphatidylinositol-4,5-Bisphosphate 3-Kinase Catalytic Subunit Alpha</i>
PIK3CB	<i>Phosphatidylinositol 4,5-Bisphosphate 3-Kinase Catalytic Subunit Beta</i>
PIK3R1	<i>Phosphoinositide-3-Kinase Regulatory Subunit 1</i>
PR	<i>Progesterone Receptors</i>
RNA	<i>Ribonucleic Acid</i>
SAC	<i>Spindle Assembly Checkpoint</i>
ShRNA	<i>Small Hairpin RNA</i>
STRING	<i>Search tool for the retrieval of interacting genes</i>

TFAP2C	<i>Transcription Factor AP-2 Gamma</i>
TFs	<i>Transcription Factors</i>
TNBC	<i>Triple-Negative Breast Cancer</i>
USF1	<i>Upstream Stimulatory Factor 1</i>
UTR	<i>Untranslated Region</i>

ABSTRACT

The purpose of our study was to identify key common genes associated with breast cancer and its diagnosis. The data from the three profiles were downloaded from the GEO database and analyzed using several bioinformatics tools. The common differentially expressed genes (DEGs) were determined using the *Venn diagram* visual presentation. *Gene Ontology* (GO) and *Kyoto Encyclopedia of Genes and Genomes* (KEGG) analyses were performed via the *DAVID* site, and protein-protein interaction networks (PPI) were built via the *STRING* database and visualized with *Cytoscape* software. Then, we checked the overall survival and expression of key genes using the *GEPIA2* database. 663 DEGs were obtained, of which 500 genes were up-regulated and 163 were down-regulated. According to GO and KEGG analyses these DEGs were mainly enriched in positive gene regulation, the phosphatidylinositol 3kinase/protein kinase B pathway, angiogenesis, cell division, mitotic spindle assembly checkpoint signaling, and mitotic sister chromatid segregation. In addition, eight hub genes were selected, one of which was associated with decreased overall patient survival and was significantly expressed in cancer tissue relative to normal tissue. Finally, analysis using Network Analyst revealed that UBE2C's regulation involves crucial interactions with eleven miRNAs and seven transcription factors, providing insights into its complex coregulatory network. In summary, the gene UBE2C (Ubiquitin Conjugating Enzyme E2 C) can be an excellent biomarker for breast cancer diagnosis and targeted gene therapy.

Keywords: breast cancer, bioinformatics, common differentially expressed genes, cancer development.

RESUME

Le but de notre étude était d'identifier les gènes clés communs associés au cancer du sein et à son diagnostic. Les données des trois profils ont été téléchargées à partir de la base de données GEO et analysées à l'aide de plusieurs outils bio-informatiques. Les gènes à expression différentielle communs (DEGs) ont été déterminés à l'aide de la présentation visuelle *Venn diagram*. Des analyses *Gene Ontology* (GO) et *Kyoto Encyclopedia of Genes and Genomes* (KEGG) ont été effectuées via le site *DAVID*, et des réseaux d'interaction protéine-protéine (PPI) ont été construits via le site *STRING* et visualisés avec le logiciel *Cytoscape*. Ensuite, nous avons vérifié la survie globale et l'expression des gènes clés à l'aide de la base de données *GEPIA2*. On a obtenu 663 DEGs, dont 500 gènes surexprimés et 163 sous-exprimés. Les analyses GO et KEGG ont montré que ces DEGs ont été enrichis dans la régulation positive de l'expression des gènes, la régulation positive de la transduction du signal phosphatidylinositol 3 kinase/protéine kinase B, l'angiogenèse, la division cellulaire, la signalisation du point de contrôle de l'assemblage du fuseau mitotique et la ségrégation des chromatides sœurs mitotiques. De plus, huit gènes clés ont été sélectionnés, dont un était associé à une diminution de la survie globale des patients et était significativement exprimé dans les tissus cancéreux par rapport aux tissus normaux. Enfin, l'analyse utilisant la base de données *NetworkAnalyst* a révélé que la régulation d'UBE2C implique des interactions cruciales avec onze miRNAs et sept facteurs de transcription, fournissant des informations sur son réseau de corégulation complexe. En résumé, le gène UBE2C (Ubiquitin Conjugating Enzyme E2 C) peut être un excellent biomarqueur pour le diagnostic du cancer du sein et la thérapie génique ciblée.

Mots clés : cancer du sein, bio-informatique, les gènes à expression différentielle communs, développement du cancer.

الملخص

كان الهدف من دراستنا هو تحديد الجينات المشتركة المحورية المرتبطة بسرطان الثدي وتشخيصه. استُخرجت البيانات من ثلاث مجموعات مختلفة من قاعدة البيانات *GEO*، وخضعت لتحليل متقدم باستخدام عدد من أدوات المعلوماتية الحيوية. جرى تحديد الجينات مختلفة التعبير (*DEGs*) باستخدام المخطط المرئي *venn diagram*، فيما أُجريت تحليلات التأثير الجيني (*GO*) ومسارات (*KEGG*) *Kyoto Encyclopedia of Genes and Genomes* عبر منصة *DAVID*. كما أنشئت شبكات التفاعل البروتيني (*PPI*) بالاعتماد على قاعدة *STRING*، وتم تصويرها باستخدام برنامج *Cytoscape*. بعد ذلك، لتحقيق فهم أعمق لأهمية هذه الجينات، أُجري تحليل معدل البقاء الكلي والتعبير الجيني للجينات الرئيسية باستخدام قاعدة البيانات *GEPIA2*. بلغ عدد الجينات مختلفة التعبير 663 جيناً، منها 500 مفرطة التعبير، و163 منخفضة التعبير. وقد أظهرت نتائج تحليل *GO* و *KEGG* أن هذه الجينات تشارك في تنظيم التعبير الجيني الإيجابي، مسار فوسفاتيديل اينوزيتول 3 كيناز/ بروتين كيناز B، تكوّن الأوعية الدموية، انقسام الخلية، مراقبة تجمع المغزل الانقسام، بالإضافة إلى فصل الكروماتيدات الشقيقة خلال الانقسام الخلوي. ومن بين الجينات التي تم تحديدها، برزت ثمان جينات رئيسية، أحدها ارتبط بشكل واضح بانخفاض في معدل البقاء الكلي للمرضى، كما أبدى تعبيراً مرتفعاً في الأنسجة السرطانية مقارنة بالنسيج الطبيعي. أخيراً، كشف تحليل *Network Analyst* أن تنظيم الجين *UBE2C* يتضمن تفاعلات مهمة مع أحد عشر *miRNAs* وسبعة عوامل نسخ، مما يوفر معلومات قيمة حول شبكته التنظيمية المعقدة. وبناءً على ما سبق، يُعد الجين *UBE2C* (Ubiquitin Conjugating Enzyme E2 C) مرشحاً واعداً ليكون مؤشراً حيويًا لتشخيص سرطان الثدي، بالإضافة إلى كونه هدفاً محتملاً للعلاج الجيني الموجه.

الكلمات المفتاحية: سرطان الثدي، المعلوماتية الحيوية، الجينات مختلفة التعبير، تطور السرطان.



INTRODUCTION

Breast cancer is the most common cancer in women and the second-leading cause of cancer-related deaths among women worldwide. Breast cancer makes up about 24.2% of all new cancer cases in women. It is expected to represent 1.7% of all cancer cases by 2025, with around 2.3 million new cases in women, surpassing lung cancer as the leading cause of cancer worldwide. A review of the literature reports that in 2018, there were about 2 million new breast cancer cases and 626,679 deaths. This cancer can also affect younger women [1].

Breast cancer, a complex condition characterized by major variance at both the genetic and clinical levels, stands out in this globally difficult situation. Usually grouped according to their spread, structure, and expression of particular immunohistochemical markers, along with current genetic analysis, most breast cancers are adenocarcinomas. Research has revealed that different traits are connected to differences in the cancer's treatment response and its probable outcomes [2]. In situ breast cancers are also limited to the ducts or lobules of the breast [3], where ductal carcinoma in situ (DCIS) is thought to be more prevalent than lobular carcinoma in situ (LCIS). Although the exact causes of the formation and advancement of in situ tumors are not completely known, lobular carcinoma in situ is not thought to have the possibility to mutate into a malignant tumor [4-5]; still, both types are considered risks for invasive breast cancer [6].

Even though breast cancer therapy has come a long way, it is still the most prevalent cancer, with the highest incidence rates among women worldwide. Thus, precise diagnostics, efficient therapy, and improved prediction of the disease's outcome would therefore depend greatly on one's knowledge of the biological pathways behind breast cancer [7].

The identification of novel biomarkers for BC is crucial to its diagnosis, therapy, and prognosis. In the last decade, the rapid development of high-throughput techniques and public gene databases enables to filter out a wider range of disease-associated genes based on abundant data by utilization of microarrays, analyze them holistically, and thus identify potential new drug targets for early diagnosis and treatment [8]. The use of bioinformatics tools to analyze data from gene expression research offers a promising field that opens new possibilities and helps to develop creative diagnostic and therapeutic approaches [9].

Our study is based on the use of multiple bioinformatics tools and databases to extract and analyze a dataset in order to identify common differentially expressed genes in breast cancer tissues. Thus, our study aimed to identify new biomarkers involved in BC progression that could be used as potential targeted therapies.

This work is divided into three parts:

- The theoretical part is specific to breast cancer: Risk factors of breast cancer and symptoms, types, stages, diagnosis, and treatment.
- The first practical part (Methods) is dedicated to the bioinformatics tools used for analysis of data from patients with breast cancer.
- The results and discussion section which shows the different data analyses and opposes them with scientific arguments.

THEORETICAL PART

CHAPTER I: Breast cancer

1. Breast cancer

Among the most commonly diagnosed cancers is BC, which ranks second as the leading cause of cancer-related deaths among women [10]. Breast cancer is when the cells in the breast grow abnormally [11].

Apart from being the most frequently diagnosed cancer in 154 out of 185 countries, BC is the leading cause of cancer-related deaths in over 100 countries. According to the American Cancer Society, around 600 men and 40,000 women lose their lives to BC each year [12].

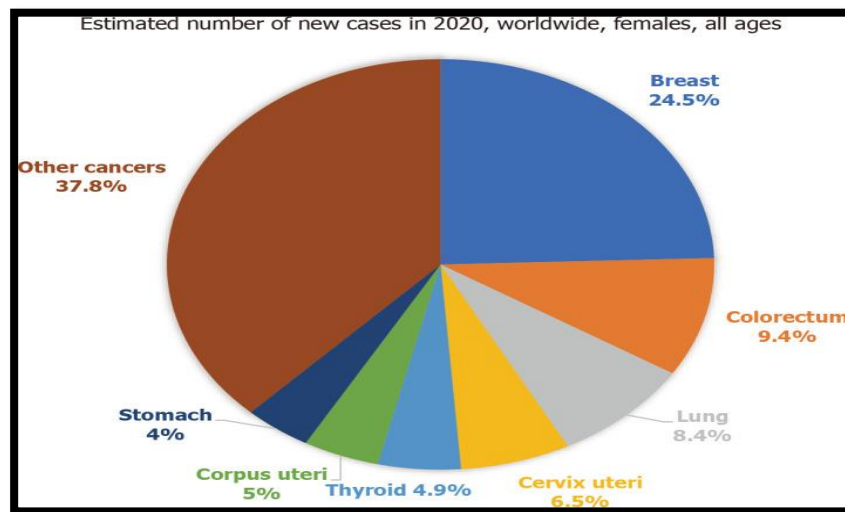


Figure 01: Estimated number of new cancer cases in 2020, worldwide [13].

2. Breast cancer risk factors

2.1. Demographic and genetic factors

- Age

Most BC patients are over 50 years old, with more than 40% being over 65 years old. The risk of getting breast cancer goes up with age: it's 1.5% at 40, 3% at 50, and over 4% at 70 [14].

- Race or ethnicity and geographic location

BC affects different races and ethnicities in different ways. The rates vary by race, and there are concerning differences in survival rates for women with breast cancer .

Black women have a higher risk of dying from breast cancer, but white women are more likely to be diagnosed with it [15].

- **Family history**

A family history of BC is an important factor strongly linked to a higher likelihood of developing BC. About 13-19% of individuals diagnosed with breast cancer mention having a first-degree relative who has the same condition. Additionally, the risk of BC increases significantly as the number of first-degree relatives affected by the disease grows [14].

- **Mutations**

Mutations in the BRCA1 and BRCA2 genes increase the risk of BC. These genes help repair DNA damage in the body, but when mutations occur, the body becomes less able to fix the damage, which increases the likelihood of cancer. Women with these mutations have a higher risk of developing breast and ovarian cancer [16].

2.2. Hormonal factors

Hormonal imbalance is one of the major causes of BC; hormones like estrogen and progesterone stimulate the growth of breast cells by binding with the ER and PR receptors. Once estrogen binds to ER or progesterone to PR, a cascade of signals inside the cell is triggered that can regulate cell growth and inhibit cell death. But if there's too much estrogen, or the signaling with progesterone is interrupted, these signals can become uncontrolled, triggering uncontrolled cell growth and tumor formation [17].

2.3. Lifestyle factors

- **Smoking**

The cancer-causing substances in tobacco smoke move through the lung alveoli into the bloodstream, where they can then be carried to the breast via lipoproteins in the plasma [18].

- **Obesity**

Obesity is linked to an increased risk of BC because it raises levels of certain hormones and receptors, like estrogen, which is associated with estrogen-positive BC. It can also cause genetic changes due to the large amount of fat tissue, leading to insulin imbalances. Additionally, obesity can trigger inflammation in the body, which may increase the chances of developing or worsening cancers, including BC [19].

- **Alcohol consumption**

Although alcohol consumption is associated with increased risk of BC, the most researched explanations include how alcohol affects estrogen levels in the body and how it interacts with receptors in breast cells, as well as the harmful effects of alcohol's breakdown products. New studies suggest that alcohol might also contribute to BC by affecting a process called epithelial-mesenchymal transition (EMT), which is involved in how cells change and spread [20].

2.4. Environmental factors

BC environmental risk factors involve contact with substances like pesticides, industrial pollutants, chemicals found in common consumer products, and physical agents such as radiation (often from X-rays) [21].

3. Symptoms

BC symptoms can vary, and some people may not have any symptoms in the early stages. Common signs include a lump or thickening in the breast or underarm; changes in the size, shape, or appearance of the breast, changes in the nipple (like it turning inward or leaking fluid), and skin changes such as, redness or the appearance of small dimples [22].

4. Breast cancer types

4.1. Molecular breast cancer subtypes:

BC is also classified into subtypes based on its molecular characteristics, such as the presence of certain hormone receptors. Understanding these patterns helps guide treatment more effectively.

There are four main patterns of molecular breast cancer subtypes:

- * Luminal A
- * Luminal B
- * HER2-positive (HER2-enriched)
- * Triple-negative [23]

4.1.1. Luminal A breast cancer

Luminal A breast cancer is characterized by the presence of hormone receptors (positive for estrogen and/or progesterone) and the absence of HER2 (negative for HER2), in addition to low levels of Ki-67, which indicates slower growth of cancer cells [23].

4.1.2. Luminal B breast cancer

Luminal B breast cancer is characterized by the presence of hormone receptors (positive for estrogen and/or progesterone) and may or may not have HER2 (HER2-positive or HER2-negative), in addition to high levels of Ki-67, which indicates faster growth of cancer cells [23].

4.1.3. HER2-enriched breast cancer

HER2-enriched breast cancer is characterized by the presence of a large amount of HER2 protein on the surface of cancer cells. This subtype is typically ER-negative and PR-negative and accounts for about 10-15% of all cases [23].

4.1.4. Triple negative breast cancer

Triple-negative breast cancer (TNBC) is diagnosed when the results of the immunohistochemical (IHC) test are negative for ER, PR, and HER2 receptors, which means the absence of these receptors in the cancer cells. TNBC accounts for about 10-20% of all invasive breast cancers and is characterized by its aggressive nature [24].

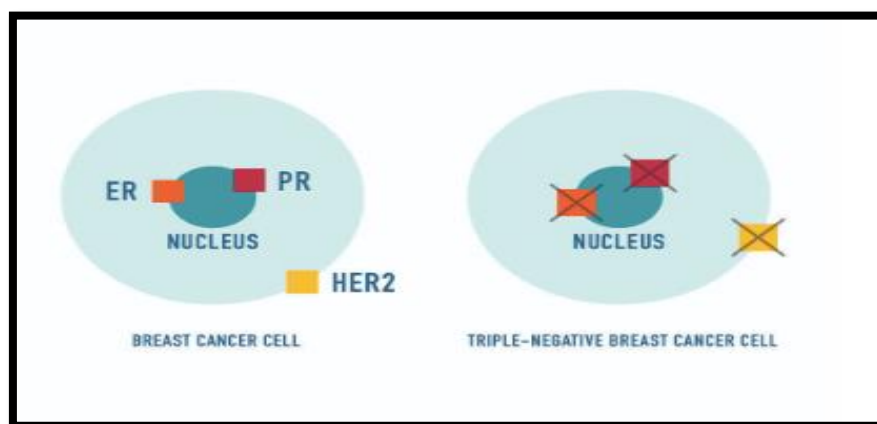


Figure 02: Illustration compares typical breast cancer cells (with ER, PR, and/or HER2) and triple-negative breast cancer cells (lacking all three) [25].

The Comparison between common molecular breast cancer subtypes is shown in Figure 03.

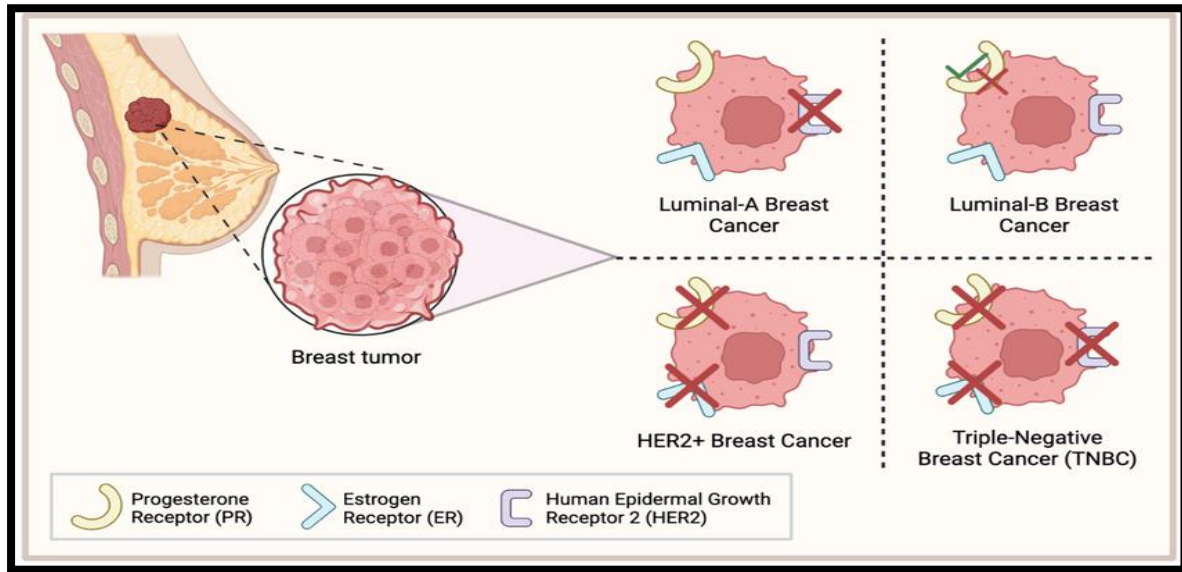


Figure 03: Classification of common molecular breast cancers subtypes [26].

5. Stages of breast cancer

✓ Stage 0:

Abnormal cells are found in the lining or parts of the breast ducts, which increases the risk of cancer developing in one or both breasts. This is a non-invasive tumor, smaller than two centimeters in size [27].

✓ Stage 1 (Early stage):

Cancer has spread to a small area of nearby tissue. This is an invasive tumor, smaller than two centimeters in size [27].

✓ Stage 2 (Localized):

The tumor measures between 2-5 cm, and some lymph nodes are affected. In some cases, the tumor is larger than 5 cm, with no lymph node involvement [27].

✓ Stage 3 (Regional spread):

The tumor exceeds 5 cm in size, with increased involvement of lymph nodes over a larger area. In some cases, no tumor may be present. The cancer could have spread to the chest wall and skin [27].

✓ **Stage 4 (Distant spread):**

Cancer has moved from the breast to other parts of the body nearby [27].

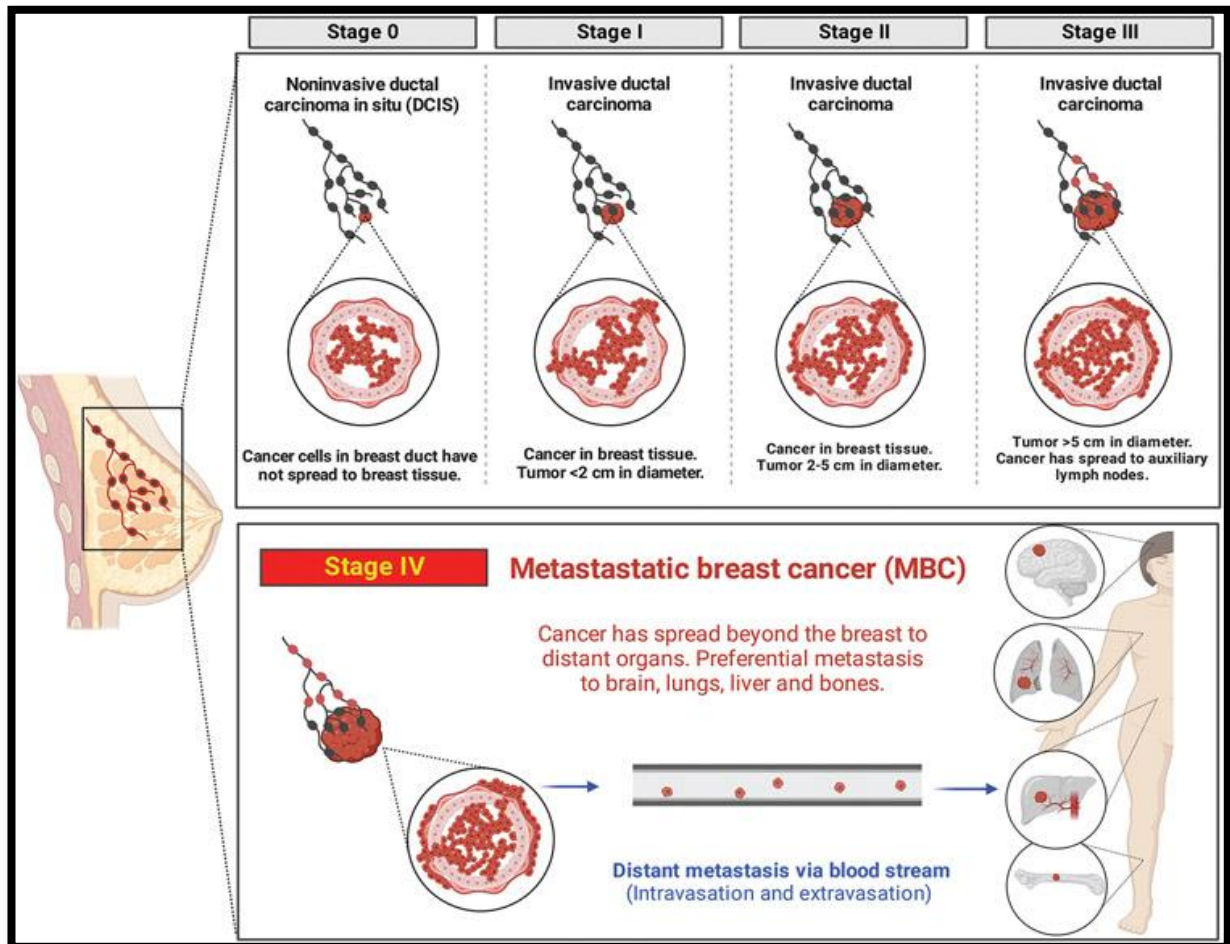


Figure 04: Illustrations showing the mechanisms of different stages of breast cancer, tumor size at each stage, 5-year survival rates, and available treatment options for breast cancer [28].

6. Methods of breast cancer screening

6.1. Clinical breast examination

Table 01: Comparison of common screening techniques [29].

Tech	Current recommendations	Advantages	Disadvantages
MAM	Recommended for women who have reached the initial age for BC screening	Convenient; economical; sensitive to microcalcification	Insensitive to high density breasts and deep lesions
DBT	Recommended, particularly for women with dense breasts, replacing DM	3D imaging, reducing tissue overlay	Greater radiation dose, examination time and cost
CEM	Recommended for women at high risk	Vascular functional imaging	Use of contrast agents; greater radiation dose
Ultra-sound	Recommended, particularly for women with dense breasts, and pregnant and lactating women	Noninvasive; real-time; no radiation; elastography; identification of cystic and solid masses	Dependent on technologist experience; insensitive to lesions without clear mass
ABUS	Recommended, particularly for women with dense breasts, and pregnant and lactating women	Less dependent on technologist skill; reproducible	Unable to assess the axillary lymph node status
MRI	Recommended for women at high risk	Most precise identification of soft tissue; reflects both the anatomical structure and lesions; displays small lesions, multifocal lesions.	Insensitive to calcification; expensive; long examination time; use of contrast agents.

6.2. Physical Examination

6.2.1. Breast self-examination (BSE)

Numerous awareness campaigns urge women to perform monthly self-examinations of the breast. However, the evidence linking breast self-exams to a reduction in mortality is scarce [30].

BSE can be performed in different ways:

- **Visual Inspection:** The breast should be examined in front of a mirror while standing and lying down, with the arm on the same side lifted. The goal is to look for any changes such as asymmetry, dimpling, inversion of the nipple, or redness.
- **Palpation:** Using the pads of the three middle fingers, gently press on the breast in a firm but circular motion, systematically checking the entire breast and underarm area.
- **Checking for Nipple Discharge:** Gently squeeze the nipple to check for any unusual discharge [31].

7. Breast cancer therapy

There are many different treatment options, and this depends on the individual, as well as the type and stage of cancer. Adjuvant therapy is when it's used after surgery and there's an absence of signs indicating cancer recurrence. Neoadjuvant therapy is when it's used before surgery to shrink the tumor and make it easier to remove, or to see how the tumor responds to a certain type of treatment. Other treatment types include:

- **Surgery:** A surgical procedure to remove the cancerous tumor.
- **Radiation therapy:** The use of high-energy X-rays to kill cancer cells.
- **Chemotherapy:** The use of powerful drugs to kill cancer cells.
- **Hormone therapy:** The use of hormones or anti-hormonal drugs to slow down or stop the growth of cancer cells, particularly in hormone-dependent cancers.
- **Targeted therapy:** The use of drugs that target specific molecules found in cancer cells, minimizing damage to healthy cells [32].

Table 02: breast cancer therapy [33].

Treatment	What Is It?	How Is It Given?	What Makes It Unique?	Common Side Effects
Surgery	Operative procedure to remove the cancer tumor from the breast	n/a	It can be performed as the only treatment, or prior to or after chemotherapy	Seroma, hematoma, lymphedema, pain, infection, cosmetic issues
Chemo-therapy	Drugs that kill cancer cells directly	Oral or intravenously	The drugs can stop or slow cancer growth and harm healthy cells	Fatigue, hair loss, bruising or bleeding, infection, anemia, nausea, vomiting, appetite changes, constipation
Hormone Therapy	Medication that is specific to types of BC that are promoted by hormones (Eg, estrogen and progesterone)	By mouth or injection in the skin	These drugs work by stopping hormones from fueling breast cancer growth	Hot flashes, vaginal dryness, night sweats, bone pain
Targeted Therapy	A drug that alters the behavior of a cancer cell	Intravenously or oral	It interferes with specific molecules involved in tumor growth and progression	Heart impairment, diarrhea, shortness of breath
Radiation	Special high-energy beams to damage cancer cells	By a machine called a linear accelerator	Targeted way to destroy cancer cells in the breast that may remain after surgery	Skin color changes such as pink or redness; itching, soreness; and possibly peeling

- ❖ Certain types, like triple-negative breast cancer, are especially hard to treat. Even though monotherapies treatments (targeted therapy, hormone therapy) can be effective for some patients, their effectiveness can decline over time, and some patients may become resistant to them. While there have been some positive outcomes with these treatments, they don't work well for everyone, especially for patients with advanced stage of BC.
- ❖ A promising new concept is combination therapy, where multiple treatments are used together to combat the disease. Different forms of combination therapy are already being applied in clinical settings and trials. Several new approaches to combining treatments have been introduced. As this treatment method becomes more common, advancements in technology are expected to bring even more creative solutions [34].

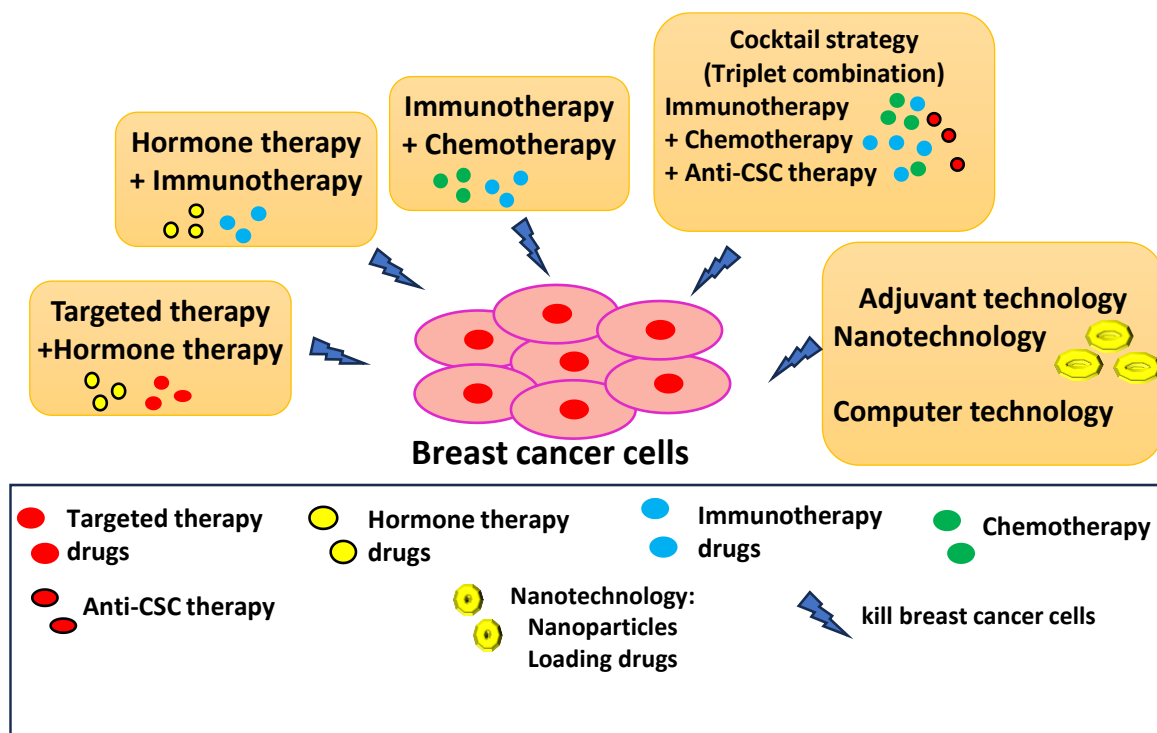


Figure 05: Schematic illustration of current treatments for breast cancer [34].



PRACTICAL PART

CHAPTER II: Methods

2. Methods

2.1. Study objective

The aim of this study was to analyze the differential expression of genes associated with BC using integrated bioinformatics tools.

2.2. Methods of analysis

2.2.1. Microarray data source and information

The microarray data used in this study were downloaded from the public functional genomics data repository, the Gene Expression Omnibus (GEO) database (<http://www.ncbi.nlm.nih.gov/gds/>). Three gene expression profiles (**GSE29431**, **GSE42568**, and **GSE61304**) of BC and normal breast tissue were chosen. The array data for GSE29431 consisted of 54 primary breast carcinomas and 12 samples of normal breast tissues, the array data for GSE42568 consisted of 104 breast cancer and 17 normal breast biopsies, the array data for GSE61304 included four RNA samples obtained from normal individuals and 58 breast tumors. All the three datasets were based on the GPL570 platform ([HG-U133_Plus_2] Affymetrix Human Genome U133 Plus 2.0 Array).

2.2.2. Data processing and identification of DEGs

The three mRNA expression microarray datasets obtained from the GEO database were analyzed using the GEO2R online tool (<https://www.ncbi.nlm.nih.gov/geo/geo2r/>).

The $|\log_2\text{-fold change (FC)}| \geq 1$ and $P \text{ value} \leq 0.05$ were conducted as the cutoff criteria for the differentially expressed genes (DEGs). Then, the raw data in TXT format were selected using Venn diagram software (<http://bioinformatics.psb.ugent.be/webtools/Venn/>) to obtain the common DEGs among the three datasets. DEGs with $\log_2\text{FC} \geq 1$ were considered up-regulated genes, while DEGs with $\log_2\text{FC} \leq -1$ were considered down-regulated genes.

2.2.3. Gene ontology and pathway enrichment analysis

Gene ontology (GO) analysis is a commonly used approach for functional studies of high-throughput transcriptome or genome data. GO and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses were performed using the online bioinformatics tool Database for Annotation, Visualization, and Integrated Discovery (DAVID) software (version 6.8, <https://david.ncifcrf.gov/>). P value ≤ 0.05 was set as the cutoff criterion.

2.2.4. PPI network and module analysis

To explore the interactions of DEGs, we used the string database (version 11.5, <https://string-db.org/>). A high confidence level (0.700) was chosen as the minimum required interaction score. After removing the disconnected nodes, protein-protein interaction (PPI) networks were visualized using Cytoscape 3.9.0 software. In addition, the Molecular Complex Detection plug-in of Cytoscape software was used to screen modules of the PPI network (degree cutoff = 2, node score cutoff = 0.2, k-core = 2, and max. depth = 100).

2.2.5. Selection of hub genes

Hub genes were extracted using the Cytoscape plug-in CytoHubba software. Then, the top eight essential hub genes were selected and ranked by Maximal Clique Centrality (MCC) and Degree topological algorithms.

2.2.6. Survival analysis and expression validation of hub genes

The online database Gene Expression Profiling Interactive Analysis GEPIA2 (<http://gepia2.cancer-pku.cn/>) was utilized to assess the effect of eight core genes on the overall survival (OS) of BC patients. To estimate the OS rate of patients with BC, samples were split into high- and low-expression groups based on the median expression. The hazard ratios (HR), corresponding 95% confidence intervals, and log-rank P values were calculated and displayed on the plot. (The GEPIA2 database tool was used to analyze the RNA sequencing expression data between BC and control samples).

2.2.7. Transcription factor-miRNA target gene

To explore the human transcription factors (TFs)-miRNA coregulatory interaction of the related key gene UBE2C, the Network Analyst database (<https://www.networkanalyst.ca/>) was used. The RegNetwork repository provided data on TF-miRNA coregulatory interactions, which facilitated the identification of regulatory TFs and miRNAs that regulate the gene UBE2C.

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PRACTICAL PART

CHAPTER III: Results and discussion

3. Results

3.1. DEGs identification

DEGs were screened among each microarray dataset using GEO2R online tool with $P \text{ value} \leq 0.05$ and $|\log\text{FC}| \geq 1$.

The GSE61304 dataset, included 1881 up-regulated genes and 1035 down-regulated genes.

The GSE42568 dataset, contained 2991 up-regulated genes and 2787 down-regulated genes.

The GSE29431 dataset, revealed 3206 up-regulated genes and 1067 down-regulated genes.

The Volcano Plots of the three GEO datasets are presented in Figure 06.

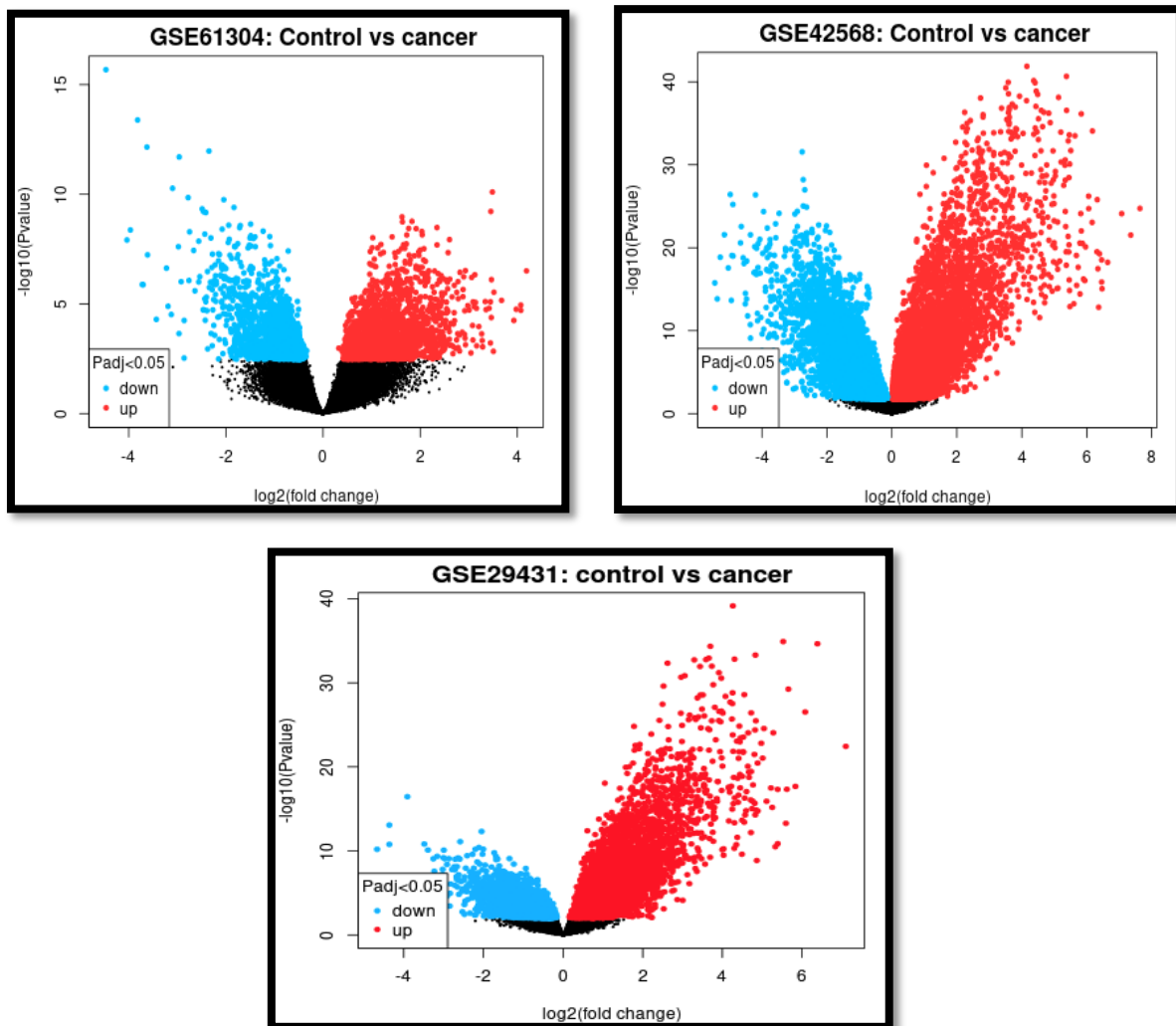


Figure 06: Volcano plots of the three GEO datasets, blue represents down-regulated genes, red represents up-regulated genes ($P \leq 0.05$ and $|\log\text{FC}| \geq 1$).

Genes were determined using a Venn diagram software of common differential expression (DEGs) between three profiles. A total of 663 common DEGs were found, including 500 up-regulated genes and 163 down-regulated genes. They were identified in all three GEO datasets, as shown in Figure 07.

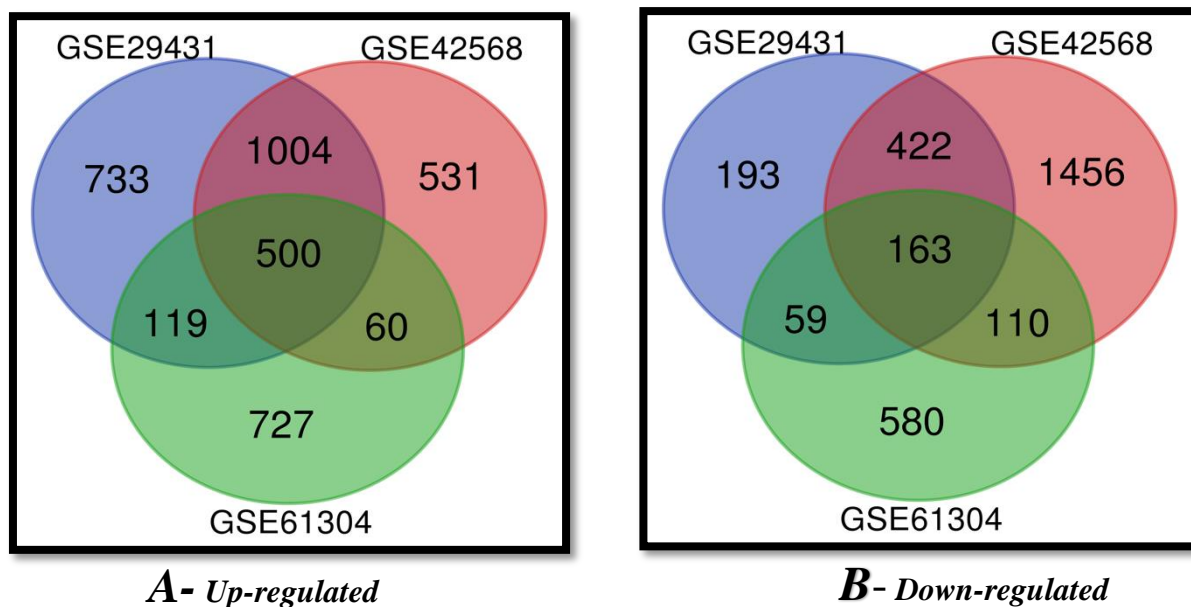


Figure 07: Identification of common DEGs in the three datasets.

A, 500 DEGs were up-regulated. B, 163 were down-regulated.

The list of common up-regulated and down-regulated DEGs from the three datasets are shown in the Table in the Annexes.

3.2. Gene ontology and pathway enrichment analysis

An enrichment analysis of GO was performed for common DEGs. The selection criterion for the analysis was $P \text{ value} \leq 0.05$. The analysis includes BP, CC and MF.

- In BP, up-regulated genes were primarily enriched in the following pathways: positive regulation of gene expression, positive regulation of phosphatidylinositol 3-kinase/protein kinase B signal transduction, angiogenesis, negative regulation of angiogenesis and response to glucose. The down-regulated genes were enriched in the following pathways: cell division, mitotic spindle assembly checkpoint signaling, mitotic sister chromatid segregation, chromosome segregation and mitotic cell cycle.
- In CC, up-regulated genes were more enriched in the following pathways: the collagen-containing extracellular matrix, caveola, extracellular region, cell surface and extracellular space. The down-regulated genes were more enriched in spindle, kinetochore, midbody, mitotic spindle and centrosome.
- In MF, up-regulated genes were extensively enriched in the following pathways: integrin binding, DNA-binding transcription factor activity, heparin binding, DNA-binding transcription activator activity, RNA polymerase II-specific and extracellular matrix structural constituent. The down-regulated genes were primarily enriched in the following pathways: microtubule binding, microtubule motor activity, protein binding, extracellular matrix structural constituent and extracellular matrix structural constituent conferring tensile strength.

The analysis of the GOs of the common DEGs is presented in Table 03.

Table 03: analysis of the GO of the common DEGs expressed in the three datasets.

Category	Term	Count	%	P-Value
Up-regulated genes				
GOTERM_BP_DIRECT	positive regulation of gene expression	36	8.0	7.5E-10
	positive regulation of phosphatidylinositol 3-kinase/protein kinase B signal transduction	19	4.2	1.9E-7
	angiogenesis	21	4.7	9.0E-7
	negative regulation of angiogenesis	14	3.1	5.2E-6
	response to glucose	10	2.2	6.5E-6
GOTERM_CC_DIRECT	collagen-containing extracellular matrix	33	7.3	4.3E-11
	caveola	14	3.1	4.2E-9
	extracellular region	87	19.3	1.2E-8
	cell surface	37	8.2	1.7E-7
	extracellular space	76	16.9	2.1E-7
GOTERM_MF_DIRECT	integrin binding	16	3.5	2.3E-6
	DNA-binding transcription factor activity	31	6.9	1.7E-5
	heparin binding	15	3.3	3.4E-5
	DNA-binding transcription activator activity, RNA polymerase II-specific	26	5.8	7.6E-5
	extracellular matrix structural constituent	11	2.4	2.7E-4
Down-regulated genes				
GOTERM_BP_DIRECT	cell division	34	22,8	5,1E-26
	mitotic spindle assembly checkpoint signaling	10	6,7	8,7E-13
	mitotic sister chromatid segregation	10	6,7	3,9E-12
	chromosome segregation	13	8,7	8,5E-12
	mitotic cell cycle	14	9,4	2,6E-11
GOTERM_CC_DIRECT	spindle	18	12,1	6,8E-16
	kinetochore	16	10,7	4,9E-13
	midbody	15	10,1	9,8E-11
	mitotic spindle	13	8,7	3,3E-10
	centrosome	23	15,4	1,5E-9
GOTERM_MF_DIRECT	microtubule binding	19	12,8	3,8E-12
	microtubule motor activity	9	6,0	1,5E-8
	protein binding	129	86,6	1,6E-6
	extracellular matrix structural constituent	9	6,0	3,8E-6
	extracellular matrix structural constituent conferring tensile strength	6	4,0	2,5E-5

3.3. Analysis of Kyoto Encyclopedia of Genes and Genomes (KEGG)

The selection criterion for the analysis of KEGG was P value ≤ 0.05 .

Up-regulated genes were primarily enriched in the following pathways: Regulation of lipolysis in adipocytes, Cytoskeleton in muscle cells, AMPK signaling pathway, PPAR signaling pathway, PI3K-Akt signaling pathway. The down-regulated genes were enriched in the following pathways: Cell cycle, Oocyte meiosis, Motor proteins, ECM-receptor interaction, Cytoskeleton in muscle cells.

Table 04: KEGG analysis of common DEGs in breast cancer.

Category	Term	Count	%	P-Value
Up-regulated genes				
KEGG_PATHWAY	Regulation of lipolysis in adipocytes	12	2.7	2.0E-7
	Cytoskeleton in muscle cells	22	4.9	3.7E-7
	AMPK signaling pathway	15	3.3	2.2E-6
	PPAR signaling pathway	11	2.4	1.9E-5
	PI3K-Akt signaling pathway	21	4.7	8.2E-4
Down-regulated genes				
KEGG_PATHWAY	Cell cycle	18	12,1	9,4E-15
	Oocyte meiosis	9	6,0	2,1E-5
	Motor proteins	10	6,7	3,9E-5
	ECM-receptor interaction	7	4,7	1,0E-4
	Cytoskeleton in muscle cells	10	6,7	1,4E-4

3.4. Protein-protein interaction analysis

To better assess the interactions between the identified common DEGs, we performed the following: Create a PPI network using the *STRING* database. A high confidence level (0.700) was chosen as the minimum required interaction score. After removing the disconnected nodes, protein-protein interaction (PPI) networks were visualized using Cytoscape 3.9.0 software.

The PPI network of common DEGs is shown in Figure 08.

The plotted PPI network is composed of 65 up-regulated genes and 43 down-regulated genes.

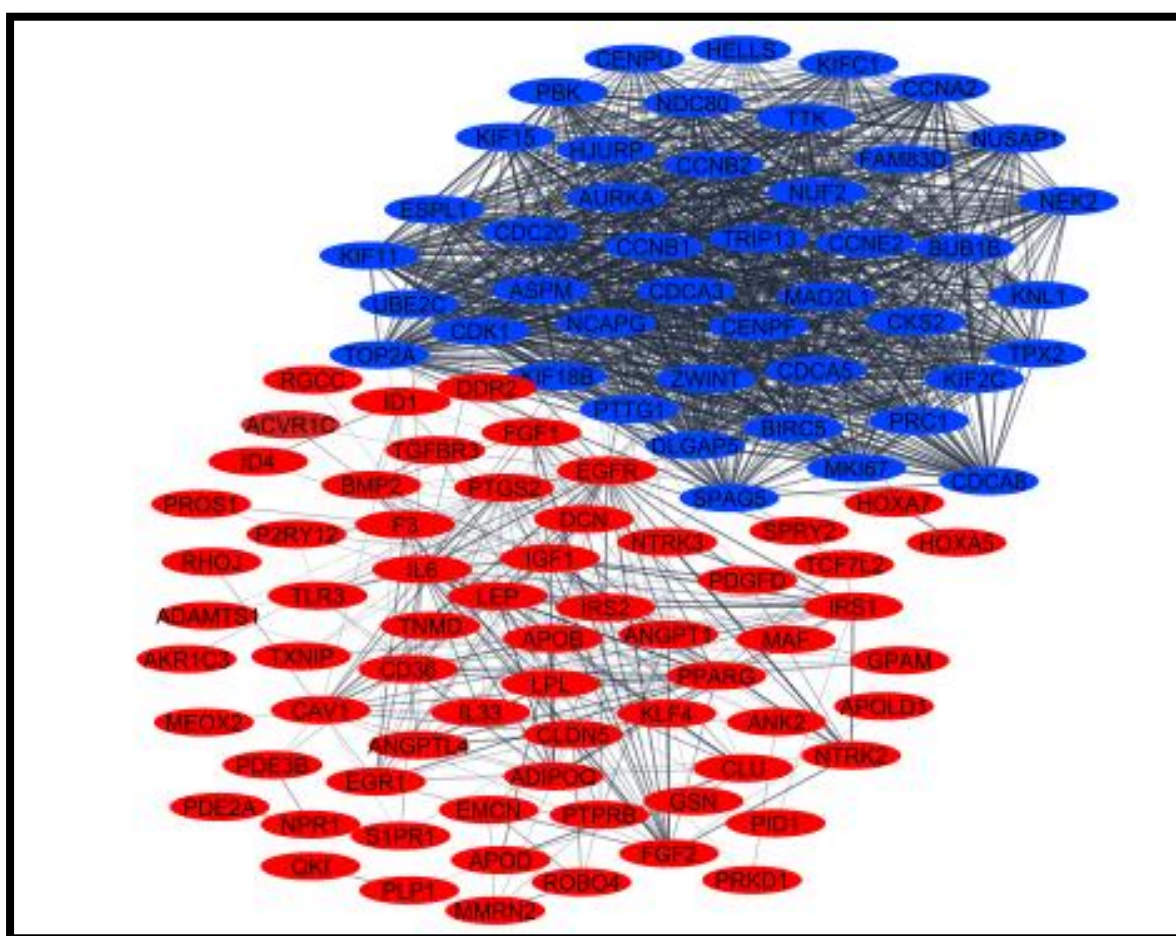


Figure 08: The PPI network created by the STRING tool and visualized by Cytoscape software (degree cutoff = 2, node score cutoff = 0.2, k-core = 2, and max. depth = 100). Red color indicates up-regulated genes; blue color shows the down-regulated genes.

3.5. Selection of hub genes

From CytoHubba, we found a functional unit of PPI network. Contained eight genes: **BUB1B**, **CDC20**, **CDK1**, **MAD2L1**, **UBE2C**, **TPX2**, **PBK**, **TOP2A**. The 8 hub genes were selected based on their high connectivity and centrality in the PPI network as determined by topological algorithms and Maximal Clique Centrality. These eight genes are considered as the hub genes associated with pathogenesis of BC (**Figure 9**).

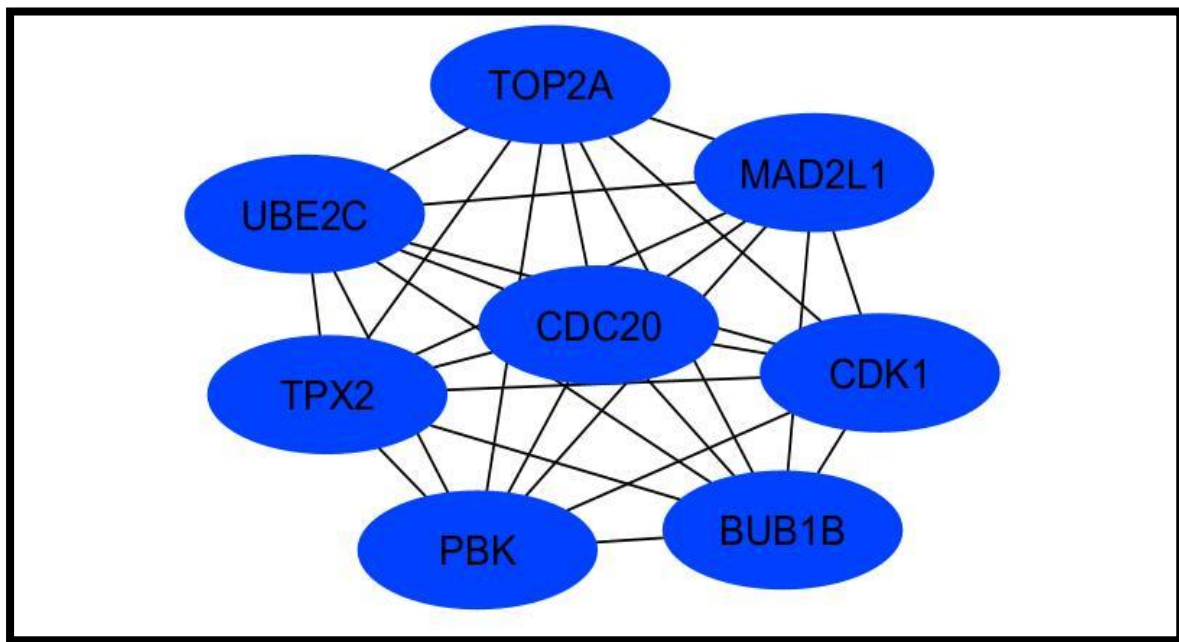


Figure 09: Top eight hub genes ranked by MCC and degree topological algorithms. All of them were down-regulated genes.

3.6. Survival analysis of hub genes

The survival analysis is used to determine the relationship between the expression profile of one or more genes and patient survival time. In this study, the overall survival of hub genes was analyzed using the GEPIA2 database to explore their prognostic value in breast cancer.

To perform the analysis, patients were divided into two groups based on the median expression value of each hub gene: a high-expression group and a low-expression group.

The log-rank test was applied as a standard method to evaluate whether these differences were statistically significant.

Each survival plot demonstrates how the expression level of a specific gene affects patient survival over time (measured in months). In these plots:

- The red line represents patients with high gene expression.
- The blue line represents patients with low gene expression.

The log-rank p-value displayed on each plot indicates whether the observed survival difference between the two groups is statistically significant:

- A p-value ≤ 0.05 suggests that the gene's expression has a significant effect on survival outcomes.

Based on the results:

- BUB1B (p = 0.26), CDK1 (p = 0.061), MAD2L (p = 0.26), CDC20 (p = 0.51), TPX2 (p = 0.11), PBK (p = 0.3), and TOP2A (p = 0.89) did not show statistically significant differences.
- However, UBE2C, with a p-value of 0.029, was the only gene among the selected hub genes that showed a significant association with poorer survival.

Therefore, these findings suggest that only the key gene UBE2C had a significantly worse survival rate

The results are illustrated in **Figure 10**.

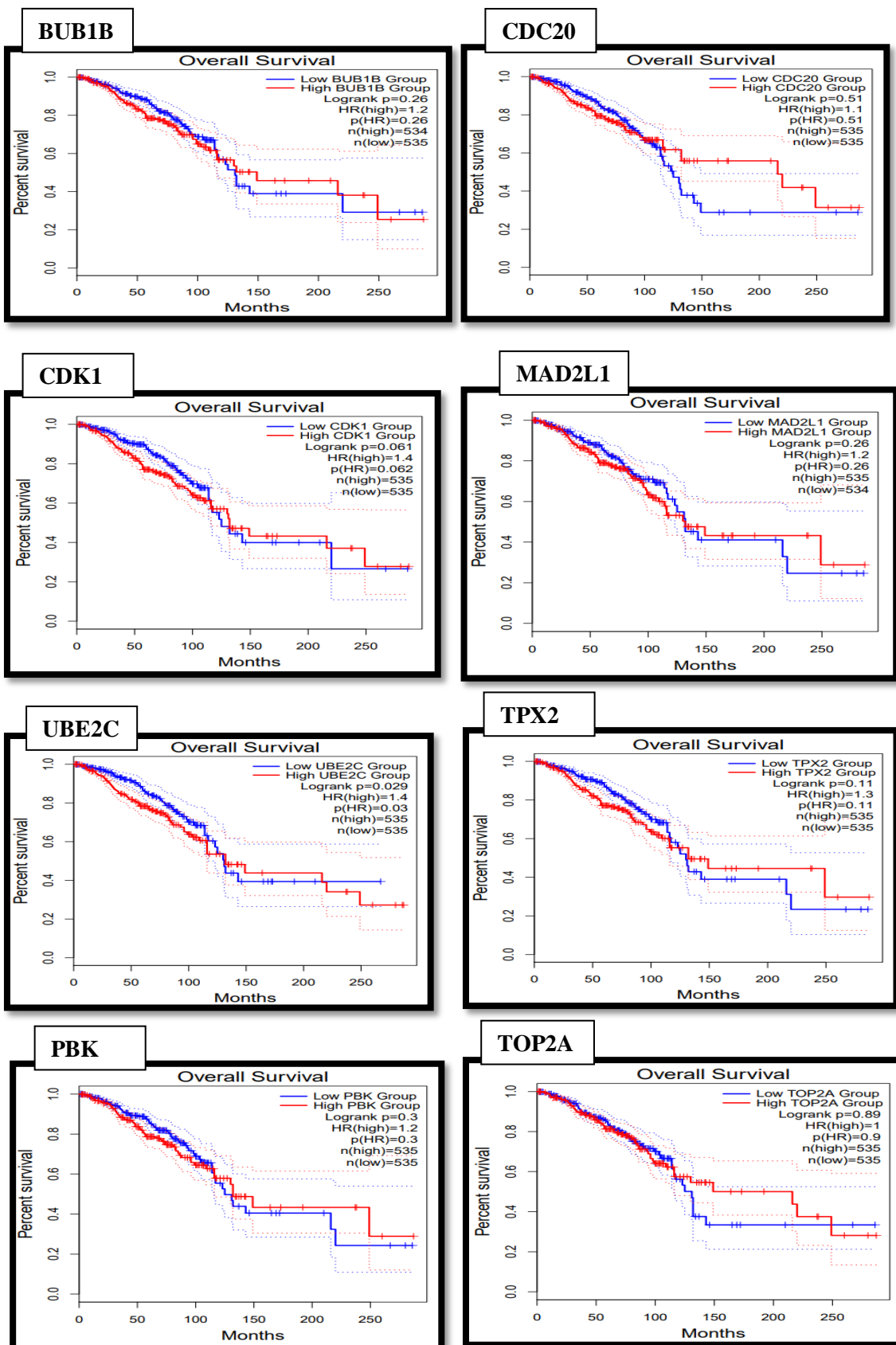


Figure 10: Analysis of overall survival of the eight hub genes by GPAIA2 database.

3.7. Validation of expression of hub genes

Then using the GEPIA2 dataset to analyze the RNA sequencing expression data between BC and control samples, each graph compares the expression of a gene between:

Tumor samples (T) shown in red and Normal samples (N) shown in gray

The red boxes are consistently higher than the gray ones, suggesting that these genes are upregulated in breast cancer tissues. The asterisk (*) above each plot usually indicates statistical significance (p value ≤ 0.05), meaning the difference in expression between tumor and normal tissues is significant. We found that the mRNA expression of the eight hub genes was higher in BC tissues than in normal breast tissues (**Figure 11**).

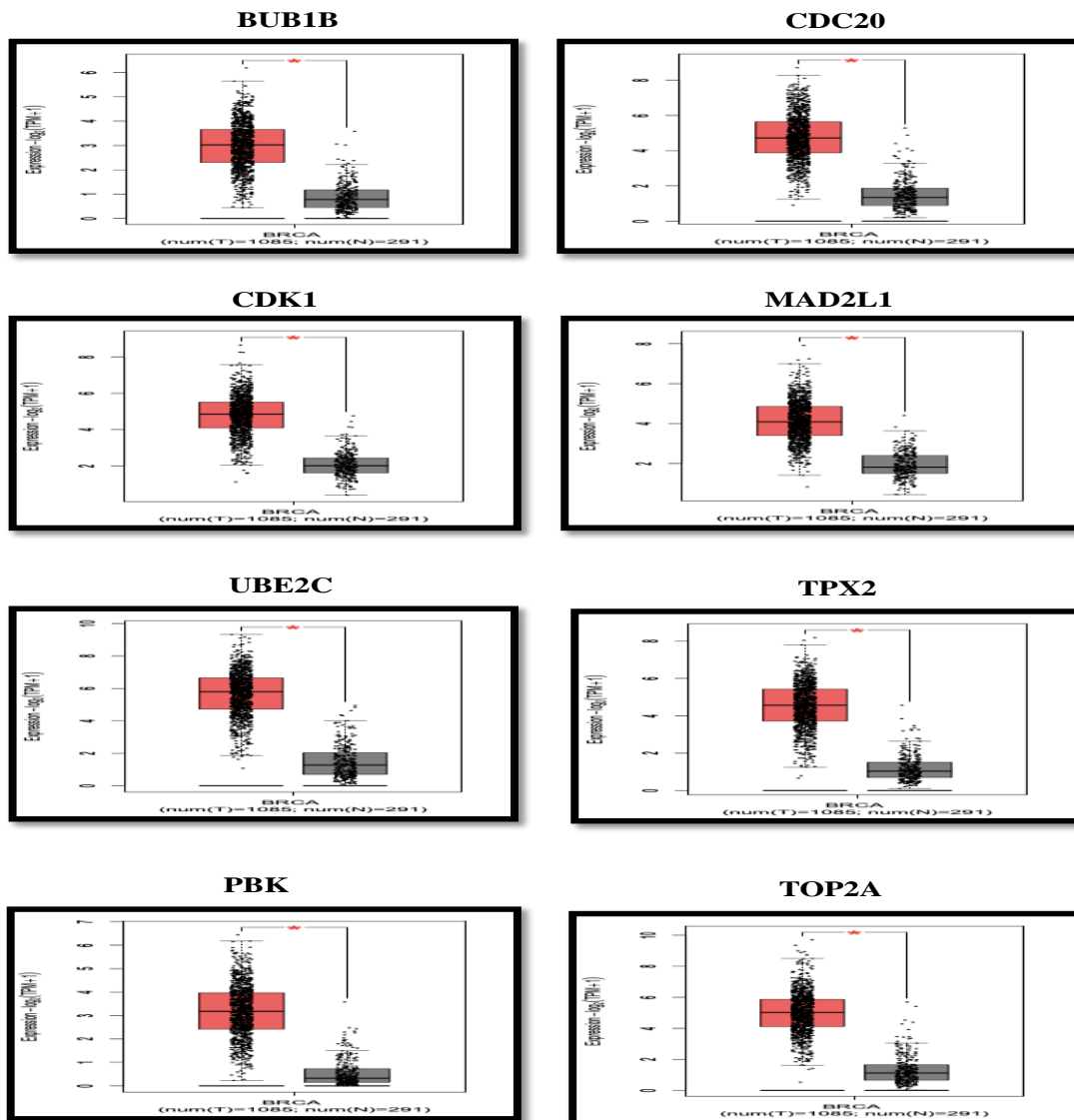


Figure 11: Expression profiles of hub genes in BC patients (GEPIA2). The red box represents BC samples; the gray box represents normal samples.

3.8. Network of transcription factors- miRNA coregulatory of key genes

Interactions between miRNAs, genes, and transcription factors (TFs) were collected using Network Analyst. The key gene UBE2C was analyzed to identify its associated miRNAs and TFs (Figure 12). UBE2C is regulated by 11 miRNAs and 07 TFs.

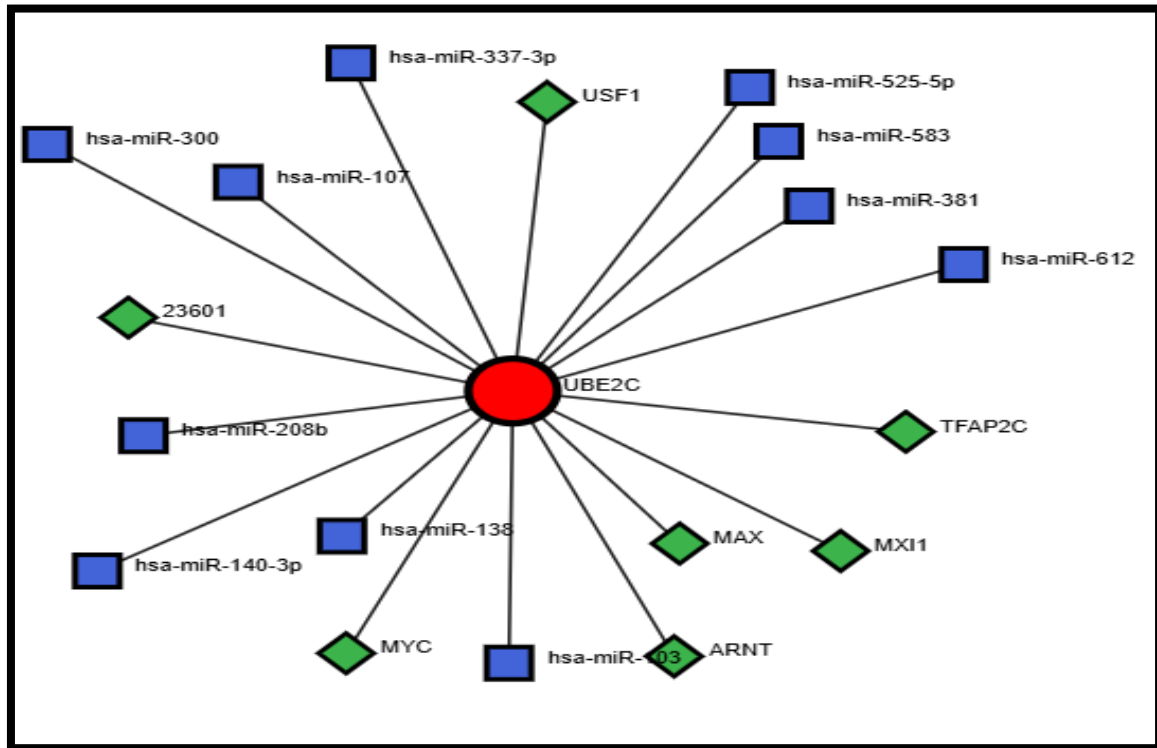


Figure 12: Network of Transcription factors- miRNA coregulatory and key gene. The red node represents the key gene (UBE2C), The blue nodes represent microRNAs (miRNAs) that interact with the key gene. The green nodes represent transcription factors.

4. Discussion

This study aims to use bioinformatics tools to analyze BC microarray data and perform comparisons to identify differently expressed genes (DEGs), we analyzed three GEO datasets (**GSE29431**, **GSE42568** and **GSE61304**) and discovered 663 overlapping differentially expressed genes (DEGs), which included 500 up-regulated genes and 163 down-regulated genes [35].

To better understand the function of these genes, the GO analysis on the common DEGs according to BP, CC and MF showed that the up-regulated genes were mainly enriched in *positive regulation of gene expression*, *positive regulation of phosphatidylinositol 3-kinase/protein kinase B signal transduction*, *angiogenesis*, *negative regulation of angiogenesis*, *response to glucose*, another level of complexity in the control of gene expression in BC is provided by microRNAs (miRNAs), small non-coding RNA molecules that bind to the 3' untranslated region (UTR) of targeted mRNAs and hence influence gene expression. Although miRNAs mostly act as negative regulators of gene expression, in particular situations, some miRNAs can also act as positive regulators. For instance, some miRNAs can prevent the translation of repressor proteins that would otherwise block their expression and therefore boost the expression of target genes. Furthermore, by indirectly affecting the expression of their target genes, miRNAs could control the activity of transcription factors. The interaction of miRNAs and transcription factors in breast cancer is a sophisticated and dynamic mechanism that helps to finely adjust gene expression patterns [36]. In addition, the phosphatidylinositol 3-kinase (PI3K)/protein kinase B (AKT) is a signaling pathway involved in cell proliferation, survival, invasion, migration, apoptosis, glucose metabolism and DNA repair [37]. The PI3K pathway is often altered in BC due to mutations or gene amplifications affecting its key components particularly the catalytic subunits p110 α (encoded by *PIK3CA*) and p110 β (*PIK3CB*), as well as the regulatory subunit p85 α (*PIK3R1*). Among these, *PIK3CA* is the most frequently mutated gene in various human cancers. It encodes the p110 α subunit and is commonly amplified in cancers of the head and neck, cervix, stomach, lungs, and breast. Notably, the highest mutation rates of *PIK3CA* have been observed in prostate, breast, endometrial, and colon cancers [38].

The link between angiogenesis and cancer is attributed to the visionary pioneer Judah Folkman (1933–2008), who was the first to propose that tumor growth is directly dependent on the development of blood vessel networks. In the case of malignant tumors, it is now clear that they have a very limited ability to grow without the support of blood vessels. Therefore,

the formation of blood vessels is a crucial step to ensure a continuous supply of essential nutrients to the tumor [39]. Studies indicate that tumors, especially BC, depend on angiogenesis, the formation of new blood vessels, for growth beyond a few centimeters. This process not only supplies essential nutrients but also facilitates tumor spread and metastasis. Tumors respond to low oxygen levels by activating angiogenesis, leading to the development of new blood vessels. This crucial process is also involved in normal functions like development and healing. Once activated, tumors enter a persistent state of angiogenesis, allowing them to enhance their blood supply and promote rapid growth [40].

On the other hand, the down-regulated genes were more enriched in *cell division*, *mitotic spindle assembly checkpoint signaling*, *mitotic sister chromatid segregation*, *chromosome segregation*, *mitotic cell cycle*. A series of distinct genetic alterations accumulates in a cell before it becomes malignant. These changes involve the activation of cellular oncogenes and the inactivation of tumor suppressor genes. Several of these various genetic changes can occur only during cell division [41]. The mitotic spindle assembly checkpoint is like a careful supervisor during cell division. It makes absolutely sure that all the chromosomes are lined up correctly and firmly attached to the spindle fibers before giving the all clear; for the cell to split. If something is wrong, this checkpoint can hit the brakes on the cell cycle. This pause is really important because it stops the cell from dividing with messed-up chromosomes, which can lead to genomic instability, a major factor in the development of cancers, including BC [42]. The SAC's command center relies on key proteins like Mad1, Mad2, BubR1, Bub3, and Mps1.

These proteins team up to form a complex that puts the brakes on the anaphase-promoting complex/cyclosome (APC/C), a vital enzyme that kicks off the chromosome separation process. When Mad2 encounters a chromosome that's not properly attached (an unattached kinetochore), it changes its shape and becomes an active blocker of APC/C. Another important player, BubR1, directly latches onto and inhibits APC/C, effectively preventing the premature pulling apart of sister chromatids [43]. These findings are in line with previous reports showing that in BC, malignant cells often exhibit defects in the spindle assembly checkpoint during mitosis, leading to inaccurate segregation of sister chromatids. This missegregation results in chromosomal instability (CIN), a hallmark of many solid tumors, including BC [44]. BRCA1, a well-known gene associated with BC susceptibility, plays a critical role in maintaining proper mitotic regulation. Loss of BRCA1 function leads to premature inactivation of the spindle checkpoint, resulting in mis segregation of sister chromatids. This contributes to the development of cells with severe genomic instability,

promoting tumor progression [45].

Then, in this study also we performed a module analysis on the PPI network constructed the eight hub genes. Through the GEPIA2 database we obtain that the genes (BUB1B, CDC20, CDK1, MAD2L1, PBK, TOP2A, TPX2, UBE2C) were significantly differentially expressed between tumor tissues and normal tissues. The OS analysis showed that the expressions of the key genes (UBE2C) may indicate that the survival chances of BC patients are low. Mitotic Arrest Deficient 2 Like 1 (MAD2L1) is a crucial member of the MAD2 protein family. Located on chromosome 4 in humans, MAD2L1 is a key component of the mitotic checkpoint complex. This complex ensures proper cell division by monitoring the process. Disrupting MAD2L1's function in mammalian cells can interfere with this checkpoint, leading to errors in cell division. Research has shown that mutations in MAD2L1 can contribute to tumor formation by causing chromosomal instability and aneuploidy (an abnormal number of chromosomes). MAD2L1 has been linked to several types of cancer, including lung adenocarcinoma, colorectal cancer, cervical cancer, hepatocellular carcinoma (HCC), acute T-cell lymphoma, BC, and stomach cancer [46].

BUB1B encodes a kinase participating in the spindle checkpoint; this protein plays a critical role in the inhibition of the anaphase-promoting complex/cyclosome (APC/C) and functions in delaying the onset of anaphase and ensuring proper chromosome segregation. The oncogene role of BUB1B has been observed in cancers such as prostate cancer, glioblastoma and gastric cancer. BUB1B is overexpressed in BC, and the level of BUB1B mRNA is significantly correlated with intrachromosomal instability. In addition, BUB1B is preferentially expressed in high-grade breast cancer, and its expression level exhibits significant associations with long-term survival [47]. BUB1B causes higher chromosomal instability in BC cells [48].

The TOP2A gene has been highlighted in eight studies as playing a significant prognostic role in BC patients. Its overexpression is associated with a poor prognosis, particularly in luminal or hormone receptor-positive BC. As a result, TOP2A could serve as both a prognostic and predictive biomarker in BC [49]. These findings align with previous studies that have shown UBE2C to be a crucial factor in cancer progression and prognosis. For example, Chao-hua Mo and colleagues examined the prognostic value of UBE2C expression at both the transcriptomic level (in 1006 cases) and the protein level (in 209 breast cancer tissue samples). Their results revealed that high UBE2C expression is linked to poorer outcomes and more aggressive tumor characteristics in breast cancer [50].

PBK has been recognized as a kinase that regulates mitosis by phosphorylating GPSM2 (G-protein signaling modulator 2) in breast cancer cells. It also interacts with p53 to control the expression of cell cycle genes. Numerous studies have found that PBK is overexpressed in various cancers, including breast, prostate, colon, bladder, and lung cancer, and serves as a prognostic biomarker for poor outcomes. Additionally, previous research has shown that PBK expression is controlled by the transcription factors Myc and E2F1 [51]. In the early stages of mitosis, CDC20 activates the anaphase-promoting complex (APC), leading to the formation of the E3 ubiquitin ligase complex APCCdc20. This complex functions to destroy key regulatory proteins of the cell cycle, thereby facilitating the process of mitosis. Additionally, CDC20 acts as a spindle checkpoint, ensuring the proper separation of chromosomes between daughter cells [52]. When CDC20 transforms into an oncoprotein, the high expression of the oncogene CDC20 has been demonstrated in various types of human malignancies [53]. Indeed, previous studies have shown that its expression significantly increases in various types of cancerous tumors, including breast, pancreatic, prostate, colon, bladder, and lung cancers. This overexpression of CDC20, whether at the gene or protein level, is associated with a poor prognosis for these tumors [54].

Cyclin-dependent kinase 1 (CDK1) (CDK1 gene) is a key enzyme that regulates the cell division cycle. It plays a vital role in the transition from the G2 phase to the M phase, where the cell prepares to divide, and also regulates the progression of the G1 phase and the transition to the S phase, where the cell grows and replicates its DNA. When there is a disruption in CDK1 activity, it can lead to uncontrolled and excessive cell growth, which is a hallmark of cancerous tumors [55]. A previous study showed that dysregulation of CDK enzymes, particularly CDK1, is associated with decreased survival rates in breast cancer patients. This study, along with our research findings, confirms that targeting these enzymes, especially CDK1, represents a promising strategy for breast cancer treatment, particularly in cases where the cancer has spread to other parts of the body (metastatic tumors) [56].

TPX2 gene, also known as XKIP2 targeting protein, plays a vital role in the regulation of microtubules associated with the cell's kinetochore. Recent studies have shown a close association between the TPX2 gene and the development of malignant tumors, including breast cancer [57]. Laboratory experiments demonstrated that inhibiting the TPX2 gene with small hairpin RNA (shRNA) hinders breast cancer cell growth and proliferation, while promoting apoptosis. This inhibition resulted in decreased PI3K expression, reduced AKT phosphorylation, and increased p53 and p21 expression, indicating that TPX2 could be a viable target for anticancer therapy [58].

Then, the interaction network highlights the role of UBE2C as a key gene regulated by 11 miRNAs and 07 transcription factors (TFs). The red node represents UBE2C, while the blue nodes correspond to miRNAs interacting with this gene. The green nodes represent transcription factors (TFs) that also regulate UBE2C expression. MicroRNAs (miRNAs) are small RNA molecules that play a role in regulating biological processes. They are processed from stem-loop regions in longer RNA transcripts. MicroRNAs are categorized into families based on their targeting characteristics, which are mainly determined by the sequence of their extended seed region. In this network, miRNAs such as hsa-miR-583, hsa-miR-337-3p, and hsa-miR-140-3p interact with UBE2C [59].

In tumor samples, UBE2C was highly expressed. When UBE2C expression was reduced, tumor cell proliferation and migration were inhibited, and the activity of the PI3K/AKT signaling pathway was decreased. Additionally, reducing UBE2C expression slowed tumor growth in animal models. The expression of UBE2C was lowered by miR-140-3p, as it directly targets UBE2C. UBE2C is overexpressed, which promotes tumor growth and migration while enhancing the activity of the PI3K/AKT pathway. However, when miR-140-3p expression was forced, these effects were partially reversed [60].

Transcription factors (TFs) play a key role in regulating various cellular processes. These processes include internal functions, like development and differentiation, as well as external ones, such as responding to signals from the environment [61]. In this network, transcription factors such as MYC, MAX, USF1 and TFAP2C regulate the UBE2C expression. TFAP2C has been implicated in the progression of breast cancer by controlling genes that are involved in regulating the cell cycle [62]. MYC is a well-known oncogene that drives cell growth and metabolism, and its interaction with UBE2C may play a key role in the progression of cancer [63]. The transcription factor MYC (MYC proto-oncogene) plays a key role as a signaling hub in various cellular processes that support the growth of many types of cancer. It regulates the expression of both protein-coding and non-coding RNAs, which in turn control essential metabolic pathways, cell death, cell proliferation, differentiation, stress responses, and drug resistance mechanisms. Activation of MYC has been commonly observed in the progression of breast cancer [64].



CONCLUSION

This study used bioinformatics tools to analyze datasets of patients with breast cancer to assess the expression levels of functional genes, key pathways, common key genes, and, subsequently, provide new insights into the prediction of the prognosis and the discovery of targeted anti-breast cancer therapies. As such, this study has allowed us to gain access to the following results:

- Identification of 663 common genes to the three expression profiles of patients, including 500 up-regulated genes and 163 down-regulated genes.
- Different biological processes are associated with breast cancer, including positive regulation of gene expression, positive regulation of phosphatidylinositol 3kinase/protein kinase B signal transduction, angiogenesis, cell division, and mitotic spindle assembly checkpoint signaling, mitotic sister chromatid segregation, chromosome segregation, and mitotic cell cycle.
- Eight genes were identified as hub genes including: BUB1B, CDC20, CDK1, MAD2L1, UBE2C, TPX2, PBK, TOP2A.
- The key gene UBE2C was identified as being significantly associated with poor survival of patients with breast cancer patients, and was confirmed to be expressed in breast cancer tissues. These findings offer key insights into breast cancer progression and the development of new anticancer therapies.



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- **Cytoscape:** is an open-source bioinformatics software platform that is used for visualizing molecular interaction networks and biological pathways: It provides a visual representation of how molecules interact with each other and the pathways they are involved in.
- **CytoHubba:** is a tool used in bioinformatics to identify hub genes or hub nodes within protein-protein interaction (PPI) networks. It is a plugin for the popular bioinformatics platform Cytoscape, which is used for visualizing molecular interaction networks.
- **DAVID (Database for Annotation, Visualization and Integrated Discovery):** is a bioinformatics resource that provides tools for understanding the biological meaning behind large lists of genes or proteins.
- **FC (Fold Change):** is a way to describe how much a quantity changes between two measurements. It is often used in biology to compare things like gene expression levels under different conditions.
- **GEO (Gene Expression Omnibus):** It is a large, public database of gene expression data housed at the National Center for Biotechnology Information (NCBI) in the United States.
- **GO (Gene Ontology):** is a major bioinformatics initiative that aims to standardize the representation of gene and protein functions across different databases and organisms. It provides a structured, controlled vocabulary of terms to describe: Molecular Function (MF), Biological Process (BP), Cellular Component (CC).
- **KEGG (Kyoto Encyclopedia of Genes and Genomes):** is a bioinformatics resource that provides a collection of databases and tools for understanding the functions of biological systems, especially at the molecular level. It focuses on pathways, genomes, diseases, and drug interactions, and is widely used in genomics, systems biology, and bioinformatics research.
- **NCBI (National Center for Biotechnology Information):** is a branch of the U.S. National Library of Medicine (NLM), part of the National Institutes of Health (NIH). It provides a wide array of databases, tools, and resources for biotechnology and biomedicine research, with a major focus on molecular biology, genetics, and health-related data.
- **OS (Overall Survival):** is a term commonly used in medical research and clinical trials, especially in oncology and other fields involving serious diseases, to measure the length

of time from either the diagnosis of a disease or the start of treatment until death from any cause. It's a key outcome used to assess the effectiveness of a treatment or intervention.

- **The p-value (probability value):** is a fundamental concept in statistical hypothesis testing, and it helps determine the significance of your results. It essentially measures the strength of evidence against the null hypothesis.
- **Venn diagram:** is a graphical representation used to show the relationships between different sets or groups. It consists of overlapping circles (or other shapes) that visually demonstrate how the sets intersect or how they are distinct from one another. Venn diagrams are often used in mathematics, logic, statistics, and even in everyday scenarios to illustrate logical relationships and set operations.
- **Volcano plot:** is a type of graphical representation used in genomics, transcriptomics, and proteomics research to visualize the results of differential expression analyses, such as gene expression studies. It combines two important statistical measures—magnitude of change (often represented by fold-change) and statistical significance (often represented by p-value)—into a single plot.
- **GEPIA2 (Gene Expression Profiling Interactive Analysis, version 2):** A web-based resource that provides interactive analysis of gene expression data derived from large-scale cancer and normal tissue datasets, such as TCGA and GTEx. GEPIA2 enables researchers to perform differential gene expression analysis, survival analysis, correlation analysis, and download gene expression data, facilitating the identification of potential biomarkers and the exploration of gene functions in cancer biology.
- **Network Analyst:** Network Analyst is a web-based platform designed for the comprehensive analysis and visualization of biological networks. It enables researchers to explore complex interactions between genes, proteins, microRNAs, and other biomolecule

Table: List of common DEGs expressed in the three datasets.

Gene symbol (up-regulated genes, 500 genes)									
CLMP	SAMD4A	IGF1	NAALAD2	PELI2	PPP1R14A	LINC01279	EPB41L4B	GNAL	RGN
LOC101930168	LOC100509620	AQP7	ERG	LEP	HOXA10-HOXA9	MIR196B	HOXA9	VGLL3	
LOC100653057	CES1	LTBP4	GHR	NAT8L	ISM1	MAOA	MRAP	EIF1	LOC101930400
MEIS2	KL	FAM162B	ID1	GPD1	NIPSNAP3B	TF	THRB	NPR1	SPTBN1
PCDH18	QKI	ACADL	SAA2	SAA1	SMIM10L2B	SMIM10L2A	FGF1	PIR-FIGF	FIGF
MYH1	ADRA1A	FHL1	RBPMS	SORBS2	KLF4	SH3D19	PLSCR4	CEBPD	ADRB1
NDRG2	SVEP1	SEMA3G	ATP8B4	GLDN	PDGFD	PREX2	HEPN1	HEPACAM	FGF14-AS2
SYNE3	LINC00341	ANKDD1A	PLAC9	SH3BGRL2	PTGS2	C1QTNF7	SPIDR	NEGR1	EHD2
RNF150	RUNX1T1	SGK2	MIR6883	PER1	S100B	BMP2	LGALS12	SEL1L2	IRS2
CLIC5	RBP7	TRHDE	DMGDH	PROS1	ENPP2	MME	LOC101930114	LOC284825	SOCS3
AMOTL2	PDK4	FILIP1	MAGI2-AS3	ST6GALNAC3	LOC101929500	CRIM1	RERGL	ANGPTL1	SLC22A3
C14orf180	EFEMP1	LINC01140	MOCS1	PLXNA4	SYN2	ADGRF5	HBA2	HBA1	ARHGAP20
LOC101926960	OGFRL1	GPIHBP1	FOXP2	NRN1	SHE	OGN	GIPC2	ANGPTL4	TLN2
NTRK3	CA4	AASS	TACC1	SPRY1	TCEAL7	LVRN	ABCA6	CXCL2	GNAI1
ARHGEF28	TNXB	TNXA	LEPROT	LEPR	MIR99AHG	TENM3	PALMD	AKAP12	MEOX2
APOLD1	LRRC70	IPO11	WNT11	ROBO4	KLB	PID1	EBF2	ZNF423	SART1
KLHL31	LHFP	HBB	GNG11	GPAM	RHOJ	SRPX	CLIP4	TFPI	ABCA9
MYZAP	TSLP	PRRG3	PLPP3	LOXL4	TGFBR3	MYL9	LRRN4CL	AKR1C1	LPL
MAF	DMD	DLC1	PPP1R1A	LINC00312	NOVA1	CCBE1	BOC	PGM5	FXYD1
RSP03	KCNIP2	DCN	KCMT2	TIMP4	SLIT2	C2orf88	HSPB7	PLP1	TMTCT1
CCDC178	SLC4A4	C2orf40	AOX1	GULP1	CFD	CH25H	RFX2	PDE3B	MOB3B
ADAMTS18	MICU3	CHRD1	LAMA2	ANXA1	NID1	ITIH5	TMEM110	MUSTN1	TMEM110
SYNPO2	PLIN4	TUSC5	CCDC50	BTNL9	MYOM1	CLDN11	MAML2	AADAC	MEOX1
ANKRD35	CACHD1	DNASE1L3	CRYAB	CD36	RNF180	DCLK1	ADAMTS9	GRK3	LIPE
PTPRB	ITGA1	ARID5B	PPARG	PQLC2L	PPP2R1B	CORO2B	MT1M	HCAR3	MID1
ZNF366	EGR1	PLAGL1	NR3C1	EMP1	PGM5-AS1	FAM149A	PKD1L2	ABCA8	SLC19A3
TMEM132C	TMEM47	CHL1	ADGRL4	HSPB2	GSN	ANK2	PLA2G4A	LMOD1	ZBTB16
EDNRB	SLIT3	MYCT1	SAA2-SAA4	SAA4	TNMD	LOC105379426	EBF3	EMX2OS	PCK1
LRRC34	VWF	ACVR1C	CIDEA	TM4SF18	CRIM1	EBF1	CD300LG	ABI3BP	RUNDC3B
ADRB2	C8orf88	G0S2	PLEKHH2	SLC7A10	NTRK3	LOC100506558	MATN2	LOC101930400	AKR1C2
SGCG	WASF3	ARRDC4	HOXA7	EGR3	TBX15	BMPER	TXNIP	ZNF521	PDE1A
TEF	TMEM100	ARHGEF40	CREB5	SAA2-SAA4	SAA2	SAA1	ACSM5	PLIN1	COPG2IT1
MFAP4	ECM2	LATS2	PDE2A	LHCGR	SCARA5	CLDN5	ATP1A2	DUSP6	IGSF10
EXOSC7	CLEC3B	ADAMTS1	NFIB	ACKR1	SORBS1	LIFR	STXBP6	DEFB132	S1PR1
LOC654342	LOC645166	NECTIN3	FOS	FGF2	CLU	PAMR1	CXCL12	PTH2R	
LOC101929583	LOC101928195	LOC100996643	LOC100133920	LOC286297	LDB2	AVPR1A	C16orf89	PRKD1	
CAV1	ALDH1A1	JAM2	TNN	PPM1L	ANKRD29	GPR146	WDFY3-AS2	GLYAT	LIPE-AS1
INMT	ZEB2	ITM2A	COX7A1	S100A10	PDE11A	KLHL29	GPRASP1	CDKN1C	AQP1
APCDD1	PCDH9	CEP126	FMO2	LMO3	SOCS2	RBMS3	CSN1S1	TMOD1	FGFBP2
ADH1B	TCF7L2	MYRIP	MIR22	MIR22HG	FABP4	KCNB1	FAM107A	CCDC85A	GPX3
ABLM3	LOC101929726	AKR1C3	DDR2	FREM1	F3	CASQ2	GGTA1P	MAMDC2	EOGT
AS1	SFRP1	GALNT16	DAAM1	COL6A6	TNS2	ZAK	C6	LOC100506718	FLRT2
Gene symbol (down-regulated genes, 163 genes)									
HIST1H3F	HIST1H3B	HIST1H3H	HIST1H3J	HIST1H3G	HIST1H3I	HIST1H3E	HIST1H3C	HIST1H3D	HIST1H3A
IST1H3A	TESMIN	TPX2	COL4A5	CCNB1	RAB3IP	CAPS	GINS1	KPNA2	COL1A1
FOXM1	CDK1	ABHD2	RGS4	CENPU	POBEC3B	SLC9A3R1	AURKA	FN1	MAD2L1
SQLE	FAM72A	FAM72D	FAM72B	FAM72C	MELK	HN1	ZWINT	CDH11	NDC80
PTTG1	CDCA5	UBE2T	CKS2	ECT2	KIF23	DEPDC1	MMP11	CCDC167	PLPP4
SPAG5	POSTN	CLDN3	CCNB2	PRC1	CDT1	CEP55	COMP	CCNE2	RALGPS2
RAD51AP1	ESRP1	CXCL10	VCAN	MKI67	SULF1	STRA13	HIST1H2BD	CERS6	COL11A1
HIST2H2AA4	HIST2H2AA3	KIAA0101	NEK2	GINS2	COL12A1	SLC44A4	ESPL1	E2F8	KIF26B
HMGB3	SAMD12	KIF11	KIF18B	KLHDC7B	IFB30	PIK3R2	KIFC1	CXCL11	SYNGR2
SLAMF8	KIF2C	GPRC5A	LRRC15	CDC20	MIAT	PLEKHF2	PBK	PRR11	TRIP13
ASPM	INHBA	CDCA3	IF16	ATAD2	KNL1	STARD10	BRIP1	UBE2C	SDC1
GJB2	RBM47	HELLS	FANCI	MSI2	CNTNAP2	KIF15	HIST1H2BC	HIST1H2BI	HIST1H2BE
HIST1H2BF	HIST1H2BG	BUB1B	HJURP	DTL	FAM83D	HMMR	AP1M2	CDCP1	HS6ST3
KIF20A	ORC6	TLCD1	COL10A1	TD02	COL5A1	TRIM59	CTHRC1	UHRF1	TTK
CDKN3	NCAPG	RM12	CENPF	NUSAP1	CDCA8				